9

Flavor of Browning Reaction Products

AHMED FAHMY MABROUK

Food Sciences Laboratory, U.S. Army Natick Research and Development Command, Natick, MA 01760

With the exception of fresh vegetables and fruits, most consumed food has to be subjected to processing (boiling, broiling, roasting, canning, baking, concentration, pasteurization,...etc.) to render it edible; to increase its consumer acceptance or to extend its shelf life. Table I lists the consumption of food commodities per capita in the United States (1). About 78% of these comodities are processed, and account for approximately 75% of the American family food budget.

Natural food flavors such as terpenes, hydrocarbons, alcohols, aldehydes, ketones, esters, acids, lactones, amines, sulfur compounds are enzymatically produced in fruits and vegetables. On the contrary, processed food develops its characteristic acceptable flavors from chemical reactions within its components at temperatures far below those at which its major components, i.e., lipids, proteins and carbohydrates pyrolyze. Food flavor precursors responsible for the productivity of volatile flavors are given in Table II.

Aqueous flavor precursors undergo nonenzymic browning during processing, which is the most important flavor producing reaction. Also, at low temperature pyrolysis, i.e., roasting between 100-270°C, these compounds undergo thermal degradation, producing new compounds some of which have desirable organoleptic notes. Some degradation products react further. For example, all purines are decomposed upon heating in acid media at temperatures above 100°C to glycine, formic acid, carbon dioxide and ammonia. Methylated purines give rise to methylamine instead of ammonia.

Proteins and glycoproteins participate in the browning reaction via their free-NH₂ groups. Furthermore, during processing, compounds of these two classes undergo hydrolysis and degradation; thus ammonia, hydrogen sulfide, peptides, amino acids, amines and sugars are produced. A delicious taste peptide which has the following structure: H-Lys-Glc-Asp-Glu-Ser-Leu-Ala-OH was isolated from beef gravy (2). In animal cells, the widely occurring peptides are carnosine, anserine, glutathione, phosphopeptides, lipopeptides and nucleopeptides. The largest class of

Table I

Consumption of Major Food Commodities Per Capita in USA, 1b

1977	154.8	93.2	3.2	1.5	56.9	12.8		34.5	44.3	9.2		16.3	4.4	289,4	17,7	54.4	4.4	11.6	3.5	17.5	21.6		82.4	25.2	57.2
1976	155.3	95.7	3.3	1.7	56.4	12.9		34.9	43.3	9.2		15.9	4.7	292.0	18.1	56.1	4.4	12.2	3.6	18.1	22.0		83.6	28.6	55.0
1975	145.4	88.9	3.5	1.8	51.2	12.2		35.4	40.3	8.6		14.5	5.0	291.1	18.7	53.3	4.8	11.2	4.0	17.3	20.3		81.3	28.7	52.6
1974	152,5	86.4	1.9	2.0	62.2	12.2		36.6	41.1	8		14.6	5.6	288.0	17.5	53.2	4.6	11.3	3.2	17.0	20.3		76.3	26.8	49.5
1970	151.4	84.1	2.4	2.9	62.0	11.8		39.5	40.5	8.0		11.5	7.1	296.0	17.7	53.0	5.3	11.0	4.7	17.3	18.2		79.1	27.9	51.2
1960	134.1	64.3	5.2	4.3	60.3	10.3		42.4	27.8	6.2		8.3	13.7	321.0	18.3	45.3	7.5	7 6	7.6	12.6	11.5		0.06	32.5	57.5
Commodity	Meats:	Beef	Veal	Lamb and mutton	Pork (excluding lard)	Fish (edibly weight)	Poultry products:	Eggs	Chicken (ready-to-cook)	Turkey (ready-to-cook)	Dairy products:	Cheese	Condensed and evaporated milk	Fluid milk and cream (prod.weight)	Ice cream (product weight)	Fats and oils-total, fat content	Butter (actual weight)	Margarine (actual weight)	Lard	Shortening	Other edible fats and oils	Fruits:	Fresh	Citrus	Noncitrus

Tal	Table I (cont'd)	(4)				
Consumption of Major For Commodity	Food Commodities Per Capita in USA, 1b 1970 1974 1975	tes Per C 1970	apita in 1974	USA, 1b 1975	1976	1977
Processed:						
Canned fruit	22.6	23.3	19.6	19.3	19.2	19.5
Canned juice	13.0	14.6	14.7	15.3	15.3	13.7
Frozen (including juices)	9.1	9.8	11.3	12.6	12.2	12.4
Chilled citrus juices	2.1	4.7	5.2	5.7	6.2	.8
Dried	3.1	2.7	2.5	3.0	2.7	2.6
Vegetables:						•
Fresh	0.96	91.0	93.6	93.9	94.5	93.1
Canned, excluding potatoes						
and sweet potatoes	43.4	51.2	53.3	52.1	52.8	52.8
Frozen, excluding potatoes	7.0	9.6	10.2	9.7	10.2	9.7
Potatoes, (including fresh						
equivalent of processed)	105.0	115.3	112.3	1.20.2	114.7	120,7
Sweetpotatoes, (Including fresh						
equivalent of processed)	6.5	5.2	5.1	5.3	5.3	5.0
Grains:						
Wheat flour	118	110	106	107	111	107
Rice	6.1	6.7	7.6	7.7	7.2	7.8
Other:						•
Coffee	11.6	10.5	9.5	0.6	9.6	6.9
Tea	9.0	0.7	0.8	0.8	0.8	ο.
Cocoa	2.9	3.1	3.0	2.6	3.0	2.7
Peanuts (shelled)	4.9	5.9	6.4	6.5	6.3	6.5
Dry edible beans	7.3	5.9	6.7	6.5	6.3	0.9
Melons	23.2	21.2	17.2	17.5	18.6	19.0
Sugar (refined)	97.4	101.8	96.6	90.2	64.7	95.7

Reference (1)

 $\label{thm:condition} \mbox{Table II}$ Food Flavor Precursors and Their Flavor Components

		Possib	le Flavor				
Precursors		Componen	ts Produced				
Class	Number	Per Compound	Per Class				
a- Aq	ueous Flavo	r Precursors					
<pre>1- Glycoproteins</pre>							
2- Glycopeptides							
3- Proteins		± _					
4- Peptides	60	40	2,400				
5- Amino Acids	35	40	1 400				
6- Amines	رڊ (40	1,400				
7- Nucleotides)						
8- Nucleotide.	}						
sugars	} 35	40	1,400				
9- Nucleotide	Į.		•				
sugaramine	- } .						
10- Nucleotide	Ş						
acetylsugarami:	ne)						
11- Peptide bound							
nucleotide	12	40	480				
12- Nucleosides	12	40	480				
13- Purines and							
pyrimidines	12	25	300				
14- Sugars)	•					
15- Sugaramine	₹ ₅₀	50	2,500				
16- Sugar phosphate	3 30	50	2,300				
17- Organic acids	25						
Subtotal	>241	>275	>8,960				
b- Non-Augeous Flavor Precursors							
l- Neutral lipids	25	50	1,250				
2- Polar Lipids	25	50	1,250				
3- Isoprenoids	150	5	750				
4- Carotenoids	10	25	250				
5- Unsaponifiable	•						
compounds							
Subtotal	>210		>3,500				
22 Grand total	>451		>12,460				

peptides in plants consist of γ -glutamyl dipeptides along with some tripeptides, one of which is homoglutathione (γ -glutamylcysteinyl- β -alanine) which replaces glutathione in some enzymic reactions (3). About thirty-five amino acids and amines occur naturally in foods. D-ribose is found in nucleotides, D-ribulose, D-lyxose, D-xylulose, glucose, fructose, sedoheptulose, and glyceraldehyde are present as phosphoric acid esters in the products of carbohydrate metabolism. Fucose is a constitutent of blood group substances which contain galactose and mannose. The presence of carbohydrate alcohols in vegetables, fruits and animals may be accounted for by a reduction of the reducing sugars, or through decarboxylation of the corresponding higher aldonic acids. Sorbose, glucose, sucrose, inositol, mannitol, pentitol and two unidentified sugars were reported in cocoa beans (4).

Sugar acids, mono-, di-, and tri-carboxylic acids, keto-, hydroxy-acids as well as unsaturated organic acids are present in foods. While uronic acids participate in the browning reaction, others might accelerate or retard the reaction.

Non-aqueous flavor precursors contribute directly and indirectly to food flavors. Boar meat lipids contain 5-androst-16-ene-3-one which is responsible for noticeable undesirable notes (5). The characteristic flavors of mutton and goat meat are attributed to the presence of odd-numbered n-fatty acids, and abnormal proportion of branched chain fatty acids $(\underline{6})$. During processing of foods, lipids undergo autoxidation, hydrolysis, dehydration, decarboxyltion, and degradation. Thermal degradation of neutral lipids, polar lipids and free fatty acids produce the following classes of compounds (7-13) :-(1)n- and iso-alkanals, (2) alkadianals, (3) oxo-alkanals, (4) alkenals, (5) alkadienals, (6) aromatic aldehydes, (7) methyl ketones, (8) saturated and unsaturated alcohols, (9) γ - and δ - lactones, (10) hydrocarbons, (11) keto- and hydroxy-acids, (12) dicarboxylic acids, and (13) saturated and unsaturated fatty acids, containing less carbon atoms than the parent ones.

Lipids in foods vary from traces as in cereals to 30-50% as in nuts. The physical state and distribution of lipids vary considerably among food items. In each item lipid distributions affect its flavor as it undergoes chemical reactions and act as a flavor components vehicle or partitioning medium. Furthermore, lipids have a pronounced effect upon the structure of food items. Fatty acids of neutral (triglycerides) and polar lipids of beef and pork are tabulated in Table III.

Pork fat contains more unsaturated fatty acids than beef and its linoleic acid content is double that in beef. Heat produced volatiles in red meat fats are listed in Table IV. The presence of pyrazines in the volatiles of beef fat is due to the presence of nitrogenous compounds in the fat amounting to 0.1-0.2%N, (Kjeldhal method). These compounds might be proteins, peptides, amino acids, and amine moities in polar lipids. Such compounds

Table III

Fatty Acids Composition of Beef and Pork Lipids

	Trig1c	erides	Polar	Lipids
	Beef	Pork	Beef	Pork
Fatty Acids	% of T	otal Fat	ty Acid	Content
Saturated				
Capric	0.1	0.1	-	-
Lauric	0.1	0.2	_	-
Myristic	2.2	1.2	2.6	2.0
Palmitic	27.5	23.9	13.2	20.0
Stearic	16.9	11.6	15.6	11.0
Total	46.8	37.0	31.4	33.0
b Monounsaturated				
Tetradecenoic	1.0	-	0.9	0.2
Palmitoleic	4.7	7.4	2.2	2.3
Oleic	41.3	45.3	21.2	16.2
Total	47.0	52.6	24.3	18.8
Dienoic				
Tetradecenoic	0.6	. –	1.3	0.6
Linoleic	4.4	8.7	20.2	27.9
Docasadienoic	-	-	-	0.9
Total	5.0	8.7	21.5	29.3
i Trienoic			_	
Linolenic	1.1	1.6	1.8	1.0
Eicosatrienoic	-	-	1.9	1.6
Total	1.1	1.6	3.7	2.6
e Tetraenoic				
Arachidonic	0.1	0.1	19.1	16.3
Total	0.1	0.1	19.1	16.3
9				

Reference (14)

Table IV
Heat Produced Carbonyls In Red Meat Fats

Volatile Compounds	Pork	Beef	Lamb
<u>AL</u> KANALS			
Ethanal(Acetaldehyde)	10	9a,10	
n-Propanal	10	9a,10	
n-Pentanal	7	8	
n-Hexanal	7,10	=	7.0
n-Heptanal	7,10	8,9,10	10
n-Octanal	, 7,10	8,9	•
n-Nonana1		8,9	10
n-Decanal	7,10	8,9	10
n-Undecanal	7 7	8,9	
n-Dodecanal	7		
METHYL-ALKANALS			
Methylpropanal		9a	•
Methylbutanal	7	8,9a	
ALKANEDIAL			
Ethanedial(Glyoxal)		9a	
OXOALKANAL			
2-0xopropanal(Pyruvaldehyde)		9a	
2-ALKENAL			
2t-Butenal(Crotonal)		9a	
2t-Heptena1	7	8,9	
2t-Octenal	7,10	8,9	
2t-Nonenal	7,10	8,9	10
2t-Decenal	7,10	8,9	10
2t-Undecenal	10	8	10
2,4-Alkadienal		•	
2,4-Heptadienal	10		
2,4-Nonadienal	10		
2,4-Decadienal	10	8,10	
2-ALKANONE			
Acetone		10	
2-Butanone		9a	
2-Decanone	7	7a	
2-Undecanone	,	7	
2-Tridecanone		8	
2-Pentadecanone		8	
2-Heptadecanone		8	

Table IV (cont'd)

Heat Produced Carbonyls in Red Meat Fats

Volatile Compounds	Pork	Beef_	Lamb	
HYDROXY-2-ALKANONE 3-Hydroxy-2-butanone(acetoin)		9a		
PYRAZINE 2,5-Dimethyl- 2-Ethyl- 2-Ethyl-3,6-dimethyl- 2-Ethyl-5-methyl- 2,3,5-Trimethyl-		8 8 8 8		

Reference (7,8,9,10) "a" designates heating in nitrogen atmosphere, otherwise in air.

degrade during heating and produce ammmonia and basic compounds. When ammonia, amino acids and degraded nitrogenous compounds react with $\alpha\text{-dicarbonyls}$ produced from lipid oxidation, pyrazines are formed.

Cooked chicken flavor concentrate (13) contained the following aldehydes; 3c-nonenal; 4c-decenal; 2t,4c-decatrienal; 2t,5c-undecadienal; 2t-dodecenal; 2t,4c-docadienal; 2t,6c- and 2t,6t-dodecadienal 2t-tridecenal; 2t,4c-tridecandienal; 2t,4c,7c-tridecatrienal; and 2t,4c-tetradecadienal. Three of these aldehydes: 4c-decenal; 2t,6c-dodecadienal; and 2t,4c,7c-tridecatrienal are typical breakdown of arachidonic acid, and to a major extent also 2t,5c-undecadienal. These aldehydes play an important role in cooked chicken flavor via the browning reaction. 2,4-Decadienal which is considered to be a key compound of chicken aroma, was found in the volatiles of cooked chicken $(\underline{15,16})$. 2,4-Heptadienal; 2,4-nonadienal and 2,4-decadienal were identified in heated pork fat (10) and heated beef fat (8,10). Dicarbonyls, dienals, trienals undergo amino-carbonyl reactions, resulting probably in the formation of meaty flavor notes in specific proportions characteristic of each species. Although their yield is small in comparison with monocarbonyls, their high reactivities are responsible for thier important role in flavor production.

The complexity of food flavor precursors is manifested by the number of compounds estimated in each class of compounds and by the possible total number in a single raw food. The possible number of flavor components produced per one compound of flavor precursor is estimated in the range 10-150, but is reduced to 5-40 to eliminate compounds that might be produced

by more than one precursor and to account for variations in processing methods. One hundred compounds were identified in thermal degradation products of glucose (17,18,19): aldehydes ketones, aromatics, furans, oxygenated furans, non-volatiles. Sugars react with ammonia at temperatures ranging from 20 to 260°C., to produce heterocyclic compounds: substituted imidazoles, substituted pyrazines, substituted piperazines, pyridine and substituted pyridines(20). At 0-120°C., α-dicarbonyls (2,3-butadione, ethanedial "glyoxal", 2-oxopropanal "pyruvaldehyde", and \alpha-hydroxycarbonyl "glycolaldehyde" react with ammonia in presence of formaldehyde to give substituted imidazoles. Dipeptides react with sugars either as an entity or as amino acids produced by hydrolysis during processing. In the first case, one compound is formed, pyrazinone (21), in the second case numerous flavor compounds are produced from the reaction of the two amino acids with sugar. The reactivity of dipeptides towards reaction with carbonyl compounds is much higher than that of amino acids. The taste of glycyl-L-leucine is very bitter, the product of its reaction with glyoxal has an astringent, a little sour and later a mild taste. Some products from the browning reaction and pyrolysis of flavor precursors react with ammonia and hydrogen sulfide, thus increasing the number of flavor components produced from food precursors. The molecular structure of the amino acid influences the flavor notes in the food. For example, thiophenes are reported in the volatiles produced from the pyrolysis of cysteine or from its reaction with glucose or pyruvaldehyde (22,24) but were absent in the case of cystine (23,24).

Hundreds of compounds have been identified in the volatile flavor components of processed foods. Hydrocarbons, alcohols, ethers, aldehydes, ketones, acids, acid anhydrides, esters, aromatic, lactones, pyrones, furans, pyridines, pyrroles, n-alkylpyrrole-2-aldehydes, pyrazines, sulfides, disulfides, thiols, thiophenes, thiazoles, trithiolanes, thialdine ...etc. Each compound has a characteristic note and a specific threshold which makes its contribution to food flavor unique. But none of the individual compounds were reported to completely produce the characteristic flavor of the processed food. This indicates that the food flavors are of mixed flavor notes of numerous compounds, while others are necessary for its synergistic effect, others exert a background of modifying effect.

Browning reaction has become a major topic of food research because of its relevance to the desirable and undesirable changes (flavor, color, texture, and nutritive value) occurring in foods during processing and storage. During the Second World War and its post era, research on food deterioration due to nonenzymic browning received considerable attention due to the problems resulting from the necessity to prepare dehydrated foods and food concentrates which had to be stored under tropical conditions without serious deterioration.

The purpose of this paper is to review the numerous papers published on flavors, tastes and odors resulting from the browning reaction. Investigations of model systems which have been observed under laboratory conditions are considered and their possible significance in basic and industrial processes will be discussed. Speculation on the possible correlation between model system results and specific processed food items will be presented. Results of recent work in our laboratory on flavor notes developed upon heating ribose with various amino acids will be discussed.

The numerous purely chemical papers are considered to be outside the scope of this paper. The reader is referred to several reviews on the subject (25-30). Comprehensive reviews on nonenzymic browning in relation to specific food problems had been published (31,32,33).

As flavor production in natural food is governed by too complicated reactions due to its complex components (Table II), chemists concentrated their research efforts on simpler systems to understand the reactions involved and their products.

Amino Acids-Sugars Model Systems

Model system studies of the reaction between single amino acids and naturally occurring substances capable of reacting with them has furnished valuable information leading toward an understanding of food flavors. The well known Strecker degradation of α -amino acids, Schonberg and Moubacher (25), has been used as the central reaction around which other amino compound degradation systems may be oriented. In this reaction $\alpha\text{-amino}$ acids are deaminated and decarboxylated by specific carbonyl compounds or others to yield aldehydes and ketones containing one carbon atom less. β -amino acids also undergo oxidative deamination and decarboxylation to a ketone with one less carbon atom; e.g., β -amino-n-butyric acid produces 2-propanone, which also results from the Strecker degradation of α -amino-isobutyric acid. Organic di- and tricarbonyls are not the only oxidants effective in Strecker degradations. Hydrogen peroxide in presence of ferrous sulfate degrade α -amino acids at room temperature; for example glycine yields formaldehyde. The co-oxidation of sulfurcontaining amino acids in an auto-oxidizing lipid system indicates that lipid peroxides act as oxidants ($\underline{34}$). The resulting carbony1 compounds from amino acids through Strecker degradation participate in the formation of desirable characteristic flavors during processing or undersirable ones responsible for flavor deterioration during storage.

Flavors and odors given by amino acids and sugars in dilute aqueous solutions at different temperatures has been the subject of intensive studies by various researchers. Tables V and VI summarize the descriptive aroma evolved from reacting carbonyl compounds with amino acids at 100, and 120/180°C., respectively

Table \

Aromas Developed by the Reactions of Carbonyl Compounds and Various Amino Compounds at 100°C

		The days	South a section of		
Amino Compounds	Dihydroxyacetone	Glucose	Fructose	Maltose	Sucrose
Glycine	baked potato (35)	caramelized sugar (36,39), faint	unpleasant carmel	weak (36)	objectionable
α-Alanine Valine	weak caramel(35) strong, yeasty	beer (40) . beer aroma (40) rye bread (37)	smell(36).	ļ	weak NH3(30)
a-Aminobutyric	hydrolyzate (35)	1101c), aromatic (3 <u>9</u>) manle(38)	i	Ţ	1
Leucine	strong, cheesy,	sweet chocolate (3Z) toast (39), bread (40)		! !	£
Isoleucine	moderate crust $(\underline{35})$	rye bread($\frac{38}{31}$) musty($\frac{37}{21}$), fruity, aromatic($\frac{39}{29}$)	1	•	i i
ser1ne	vaguely bread- 11ke (35)	maple syrup(38)	!		ł,
Threonine	very $\overline{\text{weak}(35)}$	chocolate(37)	1	i	Į Į
Phenylglycine Methlonine	baked potato (35)	bitter almond (38) overcooked sweet	objectionable	overcooked	unoleasant
		potato(36) potato(37)	chopped cabbage (36)	cabbage (36)	burned wood (36)

Table V (cont'd)

Aromas Developed by the Reactions of Carbonyl Compounds and Various Amino Compounds at 100°C

Amino Compounds	Amino Compounds Dihydroxyacetone	Glucose	Fructose	Maltose	Sucrose
Cysteine	mercaptan, H ₂ S	meat(39)	į.	į I	E E
Cystine	} }	meat, burnt	1	1	Į į
Proline	very strong cracker,crust,	curkey skin(39) corn-like(39) burnt protein		i i	· [
Hydroxyproline	toast(35) weak, vaguely like proline(35)	(37) potato (39)	4	ţ	1.
Arginine	very weak (35)	popcorn(37)	. !	Ļ	į
Histidine	very weak (35)	butterscotch(39), buttery note(39),	i i	£ ř	1
Glutamine		chocolate (37)	:	ļ	!
Methylamine Ammonia		empyreumatic taste(40) tarry odor, bitter	10)	[]	New tree
Phenylalanine	very strong, hyacinth (35)	taste(40) rancid caramel, unpleasant(36) violets(37) rose perfume(39)	stinging smell, pleasant very object sweet tionable(<u>36</u>) caramel(;	pleasant sweet caramel(<u>36</u>)	unpleasant sweet caramel(<u>36</u>)

Table V (cont'd)

Aromas Developed by the Reactions of Carbonyl Compounds ans Various Amino Commounds of 100°C

	ans var	ans various Amino Compounds at 100°C	s at 100°C		
Amino Compounds	Amino Compounds Dihydroxyacetone	Glucose	Fructose	Maltose	Sucrose
Tyrosine	I	_	1.	1	1
Aspartic	very weak(35)		!	ŀ	ţ
Glutamic	chicken broth (35)	oldwood pleasant(36)	too weak($\underline{3}6$)	too weak(36) pleasant	pleasant
Arginine	į	buttery note (39)	ł	1	car ame 1 (20)
Lysine	strong dark corn syrup(35)	baked sweet potato(36)	objectionable fried butter	unpleasant	rotten wet
α-Methylamino-		maple (38)	(36)	(36)	
α-Amino- isobutyric	!	maple(<u>3</u> 8)	1		I.

References (35,40)

Table VI

Aromas Developed by Heating Glucose with Various Amino Acids at $120^{\circ}\mathrm{C}$ and $180^{\circ}\mathrm{C}$

	Aroma De	Aroma Decordation	7,5124 - 1227.	A STATE OF THE STA	
Amino Acid	120°C	180°C	Amino Acid	Aroma Description 120°C	cription 180°C
Valine	moderate breadernet (21)	penetrating	Lysine	weak(41)	bread-11ke(<u>37</u>)
Leucine	moderate	burnt cheese (37)	Methionine	strong baked	potato(37)
Isoleucine	moderate breadcrust(41)	objectionable burnt cheese (37)	Proline	potato(41) strong bread- crust cracker	pleasant bakery aroma(37)
Threonine	weak(41)	burnt(37)	Histidine	(41) no significant	cornbread (37)
Phenylalanine	Phenylalanine strong flower	violets, lilac	Glutamine	aroma(41)	buttery note $(\frac{38}{37})$ butterscotch $(\frac{37}{37})$
Aspartic	strong bread-	$(\frac{1}{41})$ caramel($\frac{37}{3}$)	Tryptophan	$\operatorname{strong}(41)$	
Glutamic Arginine	moderate(41) no significant	brunt $sugar(37)$			
-	aroma(41)				

References (37,38,41)

(35-41). The aroma produced by the interaction of sugars and amino acids ranged from pleasant to objectionable. The molecular structure of sugar influences the resulting aroma of the products of its reaction with the amino acid. While a pleasant caramel sweet aroma developed upon reacting phenylalanine with maltose, an unpleasant caramel aroma developed with fructose, and a hyacinth aroma in case of dihydroxyacetone. While the aroma of methionine-sugar browning product was reminiscent of a pleasant. baked potato in presence of dihydroxyacetone, aroma notes of overcooked potatoes developed with glucose. In presence of a reducing disaccharide (maltose), overcooked cabbage aroma was noticed and an unpleasant burnt wood aroma developed in presence of non-reducing disaccharide (sucrose). Proline, valine and isoleucine in presence of glucose gave a pleasant bakery aroma (41). Keeney and Day (42) studied the character odor of the products of Strecker Degradation of amino acids using isatin as the oxidant (Table VII).

Table VII

Aroma of Strecker Degradation Products of α-Amino Acids with Isatin (Diketodihydroindala)

	WILL ISSUE	<u>Dikerodihydroindol</u>	<u>.e)</u>
		Sensory D	escription
Amino Acids	<u>Aldehyde</u>	Keeney & Day(42)	Others(43,44)
α -Alanine	acetaldehyde	malty	reminiscent of wine,
			coffee(43)
Valine	2-methy1-	apple	fruity, banana-like(43)
	propanal		green, pungent
	-	•	sweet(44)
Leucine	3-methy1-	malty	fruity, peach-like (43)
	butanal		burnt, green, sickly(44)
Isoleucine	2-methy1-	malty-apple	burnt, sickly (44)
	butanal		-
Norleucine	pentanal	flower	slightly fruity, herb-
		•	aceous,nut-like(43)
		•	burnt-green(44)
Pheny1-	phenylacet-	violets	hyacinth odor, resemble
alanine	aldehyde		benzaldehyde in
•			taste(43)
Glutamic		bacterial agar	-
Cystine	methional	cheesy-brothy	
References	(42,43,44)		

There is some agreement and disagreement between the sensory notes stated in Tables V and VII, which may be attributed to variations in the concentration of the reactants and lack of agreement on description of sensory notes. The discrepancies can be resolved, by a careful consideration of the specific conditions of the reaction. It is possible that both the nature and extent of the

reaction vary widely with the principal variables (concentration, temperature, heating period, specific reactants). Unfortunately, many of the papers insufficiently define the experimental conditions for an adequate evaluation of the results. The description of odor and taste notes of a product or a compound varies according to its concentration, the media used for evaluation(water, paraffin oil, skim milk...etc), and the sensory evaluator.

The data in Table VI indicate that the aromas developed when sugars and amino acids were heating at 120 and 180°C, were quite different from those produced at 100°C. Noticeable differences exist between aroma notes developed at 120°C and 180°C, this may be attributed to sugar degradation notes which become noticeable in the system heated at 135°C and above.

All aliphatic compounds containing primary or secondary amine group react in browning reaction but at different rates depending upon the molecular structure. The reactivity of the reactants (amino acids, carbonyl compounds) in the browning reaction had been reported in the literature as:

a. intensity of color developed,

b. amount of carbonyl compounds determined by GC or

as dinitrophenylhydrazones.

c. percent loss of reactants (one reactant or both). It is very hard to compare results reported in the literature, as there are various variables influencing the data. Concentration of reactants, ratio of amine compound to sugar or carbonyl, reaction in buffer or water, molarity and type of buffer used, pH of buffer, duration of reaction. Rate of carbonyl compounds formation will be the criterion used for reactivity except in few occasions where the percent loss of one of the reactants or both will be referred to in the manuscript.

Rooney et al (45) reported that the rate of carbonyl formation varied with the molecular structure of sugar. Xylose was most reactive as it produced the greatest quantity of carbonyls, followed by glucose, then maltose. In the presence of these sugars isoleucine was more reactive than phenylalanine. In a study on the Strecker degradation of valine-carbonyl, diacetyl showed the greatest reactivity followed by sorbose> arabinose>xylose>fructose>glucose>sucrose>rhamnose, Self(46). The number of volatile carbonyls produced by the reaction of glucose and glutamic at pH 5.0,6.5 and 8.0 at 100°C was almost equal. When these systems were heated at 180°C, the number of volatiles was greater and increased more drastically as pH increased to alkalinity (36). The amount of aldehydes produced from mixtures of 0.01M amino acid and 0.1M glucose in water was far below those produced in 0.01M phosphate buffer at pH6.5, which in turn was far below the amount obtained at pH 7.5. The addition of phosphate buffer increased the amount of aldehydes produced significantly. The amount of aldehyde produced is also influenced by the chain length of the acid. In general,

straingt chain amino acids produced more aldehydes than branched chain amino acids of the same number of carbon atoms (46).

Alanine, valine, serine, glutamic, glutamine, methionine, taurine, histidine, creatine, citrulline, carnosine, cystine, systeine, aspartic, asparagine, leucine, isoleucine, tyrosine, phenylalinine, tryptophan, α -aminobutyric, proline, ornithine, 2-pyrrolidone-5-carboxylic, threonine, glycine, lysine, arginine, cysteine were identified in aqueous red meat flavor precursors (47). Ribose was also reported as the major sugar. Flavor evaluation of the browning reaction products of these amino acids and ribose were undertaken. O.lM solution of each amino acid was prepared by dissolving the amino acid in 0.05M Sprensen phosphate buffer and readjusting the pH by the addition of monopotassium or dipotassium phosphate. O.lM ribose solution was prepared in the same buffer. Ten ml of amino acid and 10ml of ribose solutions were pipetted in 50ml glass ampoule and 50ml round bottom flask with 24/40 glass joint. 1.0 mMoles of insoluble amino acids were weighed directly in reaction vessels, and 10ml buffer added. To study the effect of heating on flavor of amino acids in absence of ribose, 10ml buffer were pipetted in glass ampoules and flasks along with the amino acid. The ampoules were sealed after evacuation, and heated for one hour at 180°C. The flasks were heated in glycerol bath at 100°C, after fitting with condensors. After the heating period, the flasks and the ampoules were removed, cooled in wet ice, and the contents transferred to vials and kept at -10°C. All experiments were run in triplicates. In absence of ribose, while sulfur containing amino acids solutions produced sulfury notes upon heating, none of the others developed any flavor notes. Table VIII lists the striking flavor notes produced by heating various amino acids-ribose solutions at 100° and 180°C.

Table VIII Flavor Notes Produced by Heating Amino Acid-Ribose AT 100°C and 180°C

Amino Acid	Flavor Desc			
	100°C	180°C		
Alanine	very mild caramel	sweet burnt caramelized sugar		
Valine	sickly sweet	penetrating burnt chocolate		
Serine	sweet bouillion	burnt sugar		
Glutamic	brothy, slightly sweet, lingering in mouth	roasted meat		
Methionine Cysteine Taurine	sulfury, savory sulfury, rotten egg pleasant toffee	crust of roast meat sulfury, spicy meat sickly, sweet, burnt caramel		

Table VIII (cont'd)

Flavor Notes Produced by Heating Amino Acid-Ribose At 100°C and 180°C

Amino Acid	Flavor Descri	iption 180°C
	100 0	
Tryptophan	oily aromatic, sugar sweet	oily aromatic,
Phenylalanine	sharp flower	flowery with aromatic and caramel notes, undersirable
Histidine	salty, slightly bitter caramelized toffee	pleasant slightly burnt caramel
Creatine	slightly salty	slightly sweet caramel
Tyrosine	slight caramel	custard slightly burnt sugar
Leucine	bitter almond	toasted bread
Aspartic	bread crumb	caramelized bread crust
Cystine	hard boiled egg yolk	meaty with H ₂ S note
Glutamine	caramel with burnt sugar note	butterscotch
Asparagine	desirable burnt sugar	creamy butterscotch sugar
α-Aminobutyric	undersirable burnt sugar	maple
γ-Aminobuturic	burnt sugar	maple
β-Aminobutyric	custard	map1e
Arginine	burnt sugar	buttery, burnt sugar
Ornithine	bread crumb	bread-like
Proline	bread crumb	cracker, toast
Cysteic	undersirable, sulfury	meaty, sulfury
2-Pyrrolidone -5-carboxylic	brothy	meaty, pleasant
Isoleucine	aromatic, undesirable	burnt cheese
Glycine	caramel, faint	burnt sugar
Lysine	custard	bread
Homocystine	canned milk	scorched boiled milk
Threonine	custard weak	burnt custard
Citrulline	toffee-like	meaty
Carnosine	buttery-toffee	meaty

The previous discussion covered Strecker degradation of amino acids sugar aqueous solutions. A limited number of investigations were carried out on the nature of the browning reaction at high temperature, i.e. low temperature pyrolysis and at low moisture content. Rohan and coworkers had provided an extensive research on model systems simulating cocoa nib roasting in absence of lipid. Amino acids and sugars were responsible for chocolate flavor development (48,49,50,51). While the degradation of amino acids in cocoa beans during roasting was incomplete, the degrading agent, reducing sugar was completely destroyed.

In a study on model systems prepared from single amino acid and glucose in molar ratio 2:1, approximating the composition of shell-free cocoa beans, Rohan and Stewart (52) reported that amino acid destruction from heating was temperature dependent and practically ceased after one hour. Sugar destruction contintued at a rate dependent on reaction temperature until the end of the experiment (Table IX).

Table IX

Destruction of Amino Acids and Peducing Sugars on Heating
Chocolate Aroma Precursor Extract

Heat:	ing	De	estruction Perd	cent	
°C	Min.	Amino Acids	Red. Sugars	Total Sugars	Aroma
50	30	6.2			
50	60	8.9	27.0		
50	120	8.9	41.1		
100	30	18.8	27.5	35.8	Cocoa
100	60	29.6	56.6	79.6	Cocoa
100	120	30.9	81.4	22.8	Cocoa
120	30	43.0	84.1	38.9	Cocoa
120	60	54.4	100	79.2	Cocoa
120	120	54.4	93.5	91.2	Cocoa
150	10	58.8		•	Cocoa
150	30	81.7			Cocoa
150	60	85.4			
150	120	85.1			

Reference (52)

Journal of Food Science

The failure of amino acid degradation to go to completion and the relation between maximal destruction and temperature might be attributed to the fact that individual amino acids reacted at different rates at different temperatures. For example certain amino acids, leucine, arginine, methionine, and lysine showed little or no detectable reaction at 100°C after one hour, while others were reactive at this temperature (Table X). At higher temperatures, rate of amino acid degradation was measurable and proportional to the temperature of the reaction.

The effect of increasing temperature on the amount of amino acid destroyed in one hour was noticeably greater at temperatures above 140°C than below 120°C. Between 120° and 140°C., increase in reaction temperature had less marked effect on the reaction due to changes in the physical condition of the reaction mixture. At about 150°C. glucose melts and, as it might be expected, its mixture with amino acids would begin to melt at lower temperature.

Table X

Destruction of Amino Acids After Heating With Glucose at
Different Temperatures for One Hour

Temp.			De	struct	ion Pe	rcent			
°C	A1	PhA1	Threo	G1u	Meth	Leu	Val	Arg	Lys
 80					0.9		7.0		
90		13.9	2.0	11.1	0.9		14.7	1.5	
100		21.0	22.9	23.1			23.1	1.0	4.5
110	11.3	22.2	41.2	28.8	28.5	10.5	26.2	14.5	19.5
120	24.3	23.2	42.2	26.2	40.5	19.5	28.2	29.0	28.0
130	33.4	32.0	59.4	39.0	44.3	27.2	30.2	33.8	30.2
135	33.4	5200	90.0			39.9	42.5	40.0	44.5
140	44.8	59.8	3020	50.9	51.9		·		
150	76.9	37.0		62.1			51.5		

Reference (52)

Journal of Food Science

Table XI indicates that the degree of degradation was influenced by the molecular structure of the amino acid. At 120°C the rate of destruction of amino acids by class was in the following order: sulfur-containing, hydroxy>acidic>neutral, basic, aromatic.

Table XI

Degradation of Amino Compounds After Heating for One

Hour in Presence of Glucose

Class of Amino Compound	ds Percent	Degradation	at °C	
	120	135	150	
Neutral		•		
a- n-Alkyl	20-28	. 40	45	
b- iso-alkyl		42.5		
Hydroxy	42	90		
Aromatic	23		60	
Acidic	36		51	
Sulfur-containing	41		52	
Basic	20-29	40-45	- -	
Reference (52)				

Moisture content of the reaction mixture influenced the degradation of amino acids. Rohan and Stewart $(\underline{52})$ reported that chocolate precursor aroma extracts when heated in the dry state for one hour at 100°C. , lost 30% of the amino acids by degradation. When very lightly moistened the amino acids degradation dropped to 9% under the same reaction conditions. When the reaction mixture was wetted with an equal weight of water, no amino acid degradation was noticed. Cocoa beans moisture content averaged 5 to 6% which might be sufficient to inhibit browning reaction at lower temperatures, thus in the early stages of roasting the removal of moisture is important.

Roasting cocoa beans results in the production of volatile and non-volatile compounds which contribute to the total flavor complex. 5-Methy1-2-pheny1-2-hexenal, which exhibited a deep bitter persistant cocoa note, was reported in the volatile fraction (53). It was postulated to be the result of aldol condensation of phenylacetaldehyde and isovaleraldehyde with the subsequent loss of water. The two aldehydes were the principal products of Strecker degradation products of phenylalanine and leucine, respectively. Non-volatiles contained diketopiperazines (dipeptide anhydride) which interact with theobromine and develop the typical bitterness of cocoa (54). Theobromine has a relatively stable metallic bitterness, but cocoa bitterness is rapidly noticed and disappears quickly. While cocoa bitterness is felt in the whole mouth, theobromine is recognized by the hind part of the tongue. Furthermore, cocoa bitterness is more intense than that of concentrated aqueous solution of theobromine. The following diketopiperazines had been identified as the components responsible for cocoa bitterness:- cyclo(-Pro-Leu-), cyclo(-Val-Phe-), cyclo(-Pro-Phe-), cyclo(-Pro-Gly-), cyclo(-Ala-Val-), cyclo(-Ala-Gly-), cyclo-(Ala-Phe-), cyclo(-Phe-Gly-), cyclo(-Pro-Asn-), and cyclo-(-Asn-Phe-). Those containing phenylalanine exhibited bitterness resembling that of theobromine. Diketopiperazines have stronger bitterness than the corresponding dipeptides or its two individual amino acids. They develop upon heating proteins to temperature above $100\,^{\circ}\text{C}$. Kato et al (55) reported that roasting serine at 280°C, for 30 min under slow nitrogen stream, resulted in the production of 2,5-diketo-3,6-dimethylpiperazine, which upon alkaline hydrolysis produced the dipeptide alanylalanine. bitter peptide, cyclo(-L-Leu-L-Trp-), was isolated from casein enzymic digest (56). The formation of cyclo(-Pro-Leu-) in aged sake is responsible for its bitterness (57). Recently, five bitter L-proline-containing diketopiperazines were reported in roasted malt (210°C.) used in dark beer brewing. These compounds and their approximate "bitter threshold % values" in aqueous solutions were: cyclo(-L-Phe-L-Pro-),0.1; cyclo((-L-Leu-L-Pro), 0.05; cyclo(-L-Pro-L-Pro-),0.1; cyclo(-L-Val-L-Pro-),0.1; and cyclo(-L-Ile-L-Pro-),0.05. The authors concluded that these

diketopiperazines did not contribute directly to beer bitterness and questioned their role in influencing the taste of the product (58).

More than 300 compounds had been identified in cocoa . volatiles, 10% of which were carbonyl compounds (59,60). Acetaldehyde, 2-methylpropanal, 3-methylbutanal, 2-methylbutanal, phenylacetaldhyde and propanal were products of Strecker degradation of alanine, valine, leucine, isoleucine, phenylacetaldehyde, and α-aminobutyric acid, respectively. Eckey (61) reported that raw cocoa beans contain about 50-55% fats, which consisted of palmitic (26.2%), stearic (34.4%), oleic (37.3%), and linoleic (2.1%) acids. During roasting cocoa beans these acids were oxidized and the following carbonyl compounds might be produced:- oleic: 2-propenal, butanal, valeraldehyde, hexanal, heptanal, octanal, nonanal, decanal, and 2-alkenals of C8 to C11. Linoleic : ethanal, propanal, pentanal, hexanal, 2-alkenals of C_3 to C_{10} , 2,4-alkadienals of C_9 to C_{11} , methyl ethyl ketone and hexen-1,6-dial. Carbonyl compounds play a major role in the formation of cocoa flavor components.

Another example of food in which the browning reaction occurs at near anhydrous condition is coffee. Coffee beans are usually roasted at temperatures ranging from 180° to 260°C. for specific period to obtain the desired degree of roasting (light, medium, and dark). Protein, sucrose, and chlorogenic acid were the compounds drastically destroyed in coffee beans upon roasting, Table XII "the data in this table were not corrected for dry weight loss which varied from 2 to 5%"(62).

Table XII Composition of Green and Roasted Coffee

	Percen	t, Dry Basis
Constituent	Green	Roasted
Hemicelluloses	23.0	24.0
Cellulose	12.7	13.2
Lignin	5.6	5.8
Fat	11.4	11.9
Caffeine	1.2	1.3
Sucrose	7.3	0.3
Chlorogenic acid	7.6	3.5
Protein(Based on nonalkaloid		
nitrogen)	II.6	3.1
	1.1	0.7
Reducing sugars	0.7	0.5
Unknown	14.0	31.7
Total	100.0	100.00
Reference (62)		·

Journal of Agricultural and Food Chemistry

The degree of roasting influenced the magnitude of degradation of sucrose, the major sugar present in coffee, Table XIII.

Table XIII
Effect of Roasting On Sucrose Content In Coffee
(Percent Dry Basis)

	Cold	mbian	Sant	ns
	Sucrose Content	% Loss	Sucrose Content	% Loss
Green Light Roast Medium Roast Dark Roast Reference	4.59 0.45 0.17 0.06	90.20 96.30 98.69	5.47 0.68 0.27 0.10	87.57 95.06 98.17

Feldman et al (62) found no noticeable changes in the contents of the following amino acids in roasted coffee beans proteins: alanine, glutamic, glycine, isoleucine, leucine, phenylalanine, proline, tyrosine, and valine. These findings are in agreement with those reported by Underwood and Deatherage (63). Table XIV showed the degree of destruction of amino acids in coffee bean after roasting. It is quite obvious that the magnitude of nonenzymic browning of amino acids was influenced by its molecular structure and degree of roasting.

Coffee oil contains about 47% linoleic, 8% oleic, 1% hexadecenoic, 32% palmitic, 8% stearic, and 5% behenic and longer chain fatty acids (64). As linoleic acid is the major unsaturated fatty acid in coffee oil, its major oxidation products 2,4alkadienals and hexen-1,6-dial would play a major role in volatile production. Green coffee beans contain 50 to 60% carbohydrates: 18% nonhydrolyzable cellulose, 13% hydrolyzable cellulose, 13% starches and pectins easily solublized, and 9-12% soluble carbohydrates of which sucrose is the major component. Raffinose and stachyose are the tri- and tetrasaccharides reported in robusta coffee beans. Arabinogalactan and galactomannon are the water-soluble polysaccharides reported in coffee beans ($\underline{62}$). The above mentioned carbohydrates, amino acids, lipids along with other flavor precursors produce several hundreds of volatile and nonvolatile compounds through different reactions during roasting. Molecular structures, quantities, and ratios of these compounds influence coffee flavor. The variety of coffee as well as the degree of roasting exert characteristic flavors. The molecular structure of carbonyls influence the type of pryazine formed. While refluxing rhamnose (100g) and ammonia (28%; 40m1) in water (100g) produced methyl- and ethyl-substituted pyrazines, glucose and ammonia reaction resulted in formation of methyl-substituted

Journal of Agricultural and Food Chemistry

Table XIV
Composition of Amino Acids in Green and Roasted Coffee (After Acid Hydrolysis),%

•		Haita			Columbia		Ang	Angola Robusta	ta
Amino Acid	Green	Roast I	Roast II	Green	Roast I	Roast II	Green	Roast I	Roast II
- Andread	61.1	00.0	0.00	3,61	0.00	0.00	2.28	0.00	00.00
Argunture Aggreeine	10.50	9.07	9.02	10.61	9.53	7.13	9.44	8.94	8.19
Asparagume Cysteine	3.44	0.38	0.34	2.89	0.76	0.69	3.87	0.14	0.14
Histidins	2.85	1.99	2.17	2.79	2.27	1.61	1.79	2.23	0.85
nistraine Tysine	6.19	2,54	2.74	6.81	3.46	2.76	5.36	2.23	2.56
Lysine Methionine	2.06	2.32	1.48	1.44	1.08	1.26	1.29	1.68	1.71
Cortino	5.60	1.77	1.26	5.88	2.60	0.80	4.97	0.14	00.00
Threonine	3.73	2.43	1.83	3.82	2.71	1.38	3.48	2.37	1.08
Total	39.09	20.50	18.84	37.85	22.41	15.63	32.48	17.73	14.53
Loss %		47.56	51.80		40.79	60.02		45.41	55.26

pyrazines only (53). The flavors of the products of cysteine-glucose and cysteine-pyruvaldehyde in anhydrous condition at different temperatures 80-190°C. are listed in Table XV (24) Five-mM cysteine+5-mM glucose or pyruvaldehyde reacted at different temperatures for 5 minutes and then evaluated by flavor panel.

Table XV

Flavors Produced by Cysteine-glucose And

Cyste	eine-pyruva	ldehyde at I		emperatures	
Reactants -		Flavor	Description	n, at °C.	
	80°	100°	130°	160°	190°
Cysteine & Pyruvalde- hyde	Japanese rice cracker weak.	Sesame, weak	Sesame.	Japanese rice cracker with sesame- like.	Sesame- burnt.
Cysteine	no odor	no odor	Japanese	Japanese	Sweet

rice

with

like,

sweet.

cracker

seasame-

rice

with

like.

cracker

sesame-

Reference (24)

&

Glucose

Agricultural and Biological Chemistry

sesame,

burnt.

Pyruvaldehyde is a liquid at room temperature and boils at 72°C, thus when cysteine-pyruvaldehyde mixture was heated at 80°C, the components are in solution and flavor notes reminiscent of Japanese rice cracker developed. As reaction temperatures increased gradually other flavor notes developed. In the case of cysteine-glucose system, no reaction took place until the reaction temperature reached 130°C. The flavor of cysteine-glucose was comparable to that of cysteine-pyruvaldehde at 160°C, with one exception, the glucose system had a sweet note. As temperature increased the flavor impression of both systems increased in similarity. The volatile compounds produced at 160°C in the presence of pyruvaldehyde were different from those in presence of glucose. While thiazole and thazolines were absent in the volatiles of cysteine-glucose, cysteine-pyruvaldehyde volatiles were devoid of pyridines, picolines and furans (24).

Food flavors consist of numerous compounds, none of which alone is characteristic of specific food. Classes of compounds which emcompass food flavors are:- hydrocarbons (aliphatic, alicyclic, aromatic); carbonyls (aldehydes, ketones); carboxylic acids, esters, imides, anhydrides; alcohols, phenols, ethers; alkylamines, alkylimines; aliphatic sulfur compounds (thiols, mono-, di- and tri-sulfides); nitrogen heterocyclics (pyrroles, pyrazines, pyridines); sulfur heterocyclics (thiophenes, thiazoles, trithiolane, thialidine); and oxygen-heterocyclics (lactone, pyrone, furan). Discussion will be limited to striking developments in heterocyclics.

Heterocyclic Compounds

During browning reaction in foods, heterocyclic compounds of known five- and six-membered ring systems containing one or more atoms than carbon as ring members, are produced. These compounds encompass several classes of compounds that exhibit desirable characteristic organoleptic notes, i.e. toasted, bread-like, roasted, brothy, mushroom-like, nutty, ... etc., or undesirable notes, e.g. peppery ammoniacal, obnoxious, ... etc. The nomenclatures and the molecular formulas of heterocyclic compounds that most frequently encountered in food volatiles are given in Figures 1 and 2.

Nitrogen-heterocyclic

Pyrrole and pyrrole derivatives. The chemical and biological value of pyrrole and its derivatives cannot be overemphasized, natural pigments, heme, chlorophyll, bile pigments and enzymes like cytochromes, contain pyrrole nucleus. Also, many alkaloids, proline and hydroxyproline contain the reduced pyrrole ring (pyrrolidine). Pyrrole and its derivatives are found among the products of the browning reaction products in processed foods. Alkylpyrroles have intense petroleum-like odor, but they give sweet, slightly burnt aroma on extreme dilution. On the other hand acylpyrroles have characteristic sweet smoky, and a little medicine-like odor (65). Although alkyl- and acyl- pyrroles do not exhibit favorable aroma like the desirable roasty aroma of pyrazines, they may play an important role in the characteristic roasty flavor of processed foods. In anhydrous condition, Nacetonylpyrrole, which had a bread-like aroma was isolated from the roasting products of proline-glucose, hydroxyproline-glucose, and pyrrolidine-pyruvaldehyde (66). 1-Pyrroline which exhibited an amine like or corn-like odor was the product of Strecker degradation of proline-, and ornithione-sugar or- polycarbonyl reagents in aqueous solutions (67). 2-Pyrrolealdehyde, 2-acetylpyrrole, 2-propionylpyrrole, N-methyl-2-pyrrolealdehyde, N-methyl-2-acetyl pyrrole, 5-methyl-2-pyrrolealdehyde, N-methyl-5-methyl-2-pyrrole-

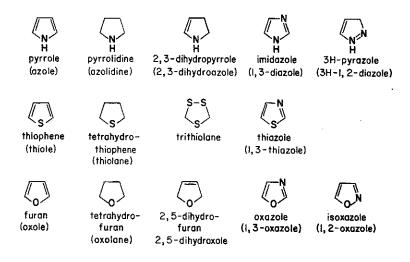


Figure 1. Five-membered ring heterocyclics

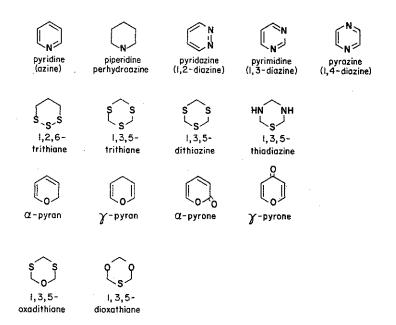


Figure 2. Six-membered ring heterocyclics

aldehyde were identified in the pyrrolic fraction of coffee volatile constituents (68, 69). N- ethylpyrrole-2-aldehyde and 5-methylpyrrole-2-aldehyde are two of the eighteen compounds reported in stored dehydrated orange juice crystals due to nonenzymic browning (70). The type of pyrrole derivatives produced depended on reaction temperature, its duration, its condition, i.e., anhydrous, aqueous, or alcoholic; pH of the media, and molecular structure of reactants. While, N-alkylpyrrole-2-aldehydes were produced upon heating D-xylose and alkylamine or amino acid in neutral aqueous or methanolic solutions at 55.100°C; the corresponding 5-methyl derivatives of N-substituted pyrrole-2-aldehydes were formed in the presence of L-rhamnose (71, 72). Odor of the products were: N-methylpyrrole-2-aldehyde (formed from D-xylose and methylamine) cinnamonaldehyde-like, N-n butylpyrrole-2-aldehyde (a product of butylamine-xylose); and 1-n-butyl-5-methylpyrrole-2-aldehyde (rhamnose and butylamine interactions product) xylene-like. 2-Formylpyrrol-1-yl-alkyl acids were isolated from the products of the browning reaction of xylose and alkylamines or amino acids in aqueous solutions (73). N-substituted-5-(hydroxymethyl)-pyrrole-2-aldehydes were formed from the reaction of the aldohexose "glucose" and alkylamines or amino acids containing primary amino group at 70 to 100°C, in neutralized aqueous, methanolic or ethanolic solutions. The resulting compounds were considerably unstable and had no odor in the pure state but developed roasted aroma with browning (74). Upon roasting glucose and several alkyl-n-amino acid (glycine, α-alanine, α-amino-n-butyric, valine, leucine, α-amino-n-caproic) at 200-250°C in two components systems; 2-5'-hydroxy-methyl-2'-formylpyrrol-1'-yl) alkanoic acid lactones were formed as the main volatile products (75). The aroma description of the prepared lactone derivatives were: propionic acid lactone (from a-alanine and glucose) caramel and a little scorching; isobutyric acid lactone (a-amino-n-butyric and glucose) maple and strong sweet; isovaleric acid lactone (valine and glucose) and isocarproic acid lactone (leucine and glucose) miso, soy sauce and a chocolate-like. The yield of propionic acid lactone after heating an equimolar mixture of 0.01 mole of glucose and a-alanine at 150, 200, and 250°C were:

Temperature, OC	Heating period, min.	Yield, unmoles
250	1	42
250	5	3 - 4
200	3	50
200	5	6
150	5	16

It is quite obvious from the above data that the propionic acid lactone disappeared during the reaction for longer periods and at higher temperatures. 2-(5'-hydroxymmethyl-2'-formylpyrrol-1'-yl) -3-methylbutanoate, and 2'-(5'-hydroxymethyl-2'-formyl-

pyrrol-1'-y1)-3-methylbutanoic acid lactone were identified in the products of the browning reaction of glucose and valine in aqueous solution at 65°C for three weeks (76). Reducing disaccharides (lactose, maltose and melibiose) reacted with alkylamine in aqueous solutions of pH 6.5, to form 1-alkyl-5-hydroxymethylpyrrole-2-aldehyde (77). N-Alkyl-2-acylpyrroles and aliphatic aldimines were the products of the reaction between furfural and its homologs with a-amino acids (78). Reactions of furfural and 2acetylfuran with glycine and valine produced a small amount of the corresponding acylalkylpyrroles which had pleasant aromas reminiscent of benzaldehyde. The resulting considerable amount of aliphatic aldimines from the reaction of furfural with valine and leucine possessed a strong odor from biting and unpleasant to mild and food-like. Nine alkylpyrroles, three acylpyrroles, three alkylpyrrole-2-aldehydes, three furfurylpyrrole, eight alkylpyrazines, and one oxazoline were identified in the volatile flavorous products of roasting DL-alanine and glucose at 250°C for one hour in nitrogen atmosphere (79). Many of the compounds identified in the reaction products of model systems had been isolated from roasted and cooked foods. For example, pyrrole, 1-methylpyrrole, 2-methylpyrrole, 1-formylpyrrole, 2-formylpyrrole, 1-formyl-2methylpyrrole, 2-formyl-1-methylpyrrole, 2-formyl-1-methylpyrrole, 2-formyl-5-methylpyrrole, 2-formyl-1-ethylpyrrole, 1-acetylpyrrole, 2-acetylpyrrole, 1-furfurylpyrrole, 2-propionylpyrrole, 2formyl-l-furfurylpyrrole were reported in roasted peanut (80). Roasted filberts volatiles contained 1-methylpyrrole, 2-pentylpyrrole, 2-isobutylpyrrole, and 2-penrylpyrridine (81).

Pyridine and its derivatives. The most unique pyridine derivative isolated from processed food is 1,4,5,6-tetrahydro-2-acetopyridine. This compound was prepared by roasting proline and dihydroxyacetone at 92°C in presence of sodium bisulfate, and exhibited a strong odor reminiscent of freshly backed soda crackers (82). 2-Ethylpyridine and 2-pentylpyridine were reported in volatile flavor components of shallow fried (83). Pyridine, 2methylpyridine, 3-methylpyridine, 2-ethylpyridine, 3-ethylpyridine, 5-ethyl-2-methylpyridine, 2-butylpyridine, 2-acetylpyridine, 2-pentylpyridine, 2-hexylpyridine, 3-pentylpyridine, 5-methyl-2pentylpyridine, and 5-ethyl-2-pentylpyridine were identified in the volatiles of roasted lamb fat (84). 2,5-Dimethylpyridine and 3,5-dimethylpyridine were tentatively identified in roasted lamb fat volatiles. The odor threshold of 2-pentylpyridine was 0.5-0.7 parts per 109 parts of water. The dilute solution of 2-pentylpyridine has a fatty or tallow-like odor. The authors attribute the unacceptance of lamb by some consumers to the high content of alkylpyridines in roasted lamb. While pyridine derivatives have burnt, heavy fruity odors (85); pyridine has a disagreeable characterisitic odor and sharp taste; and piperidine has a peppery ammoniacal odor (86). Pyridine, α-picoline (2-methylpyridine), β-picoline (4-methylpyridine), and 4-ethylpyridine were identified

in coffee aroma (87).

Pyrazines. In the thirties, the attention on pyrazines was focused on its industrial role in dyes, photographic emulsions and chemotherapy. Its importance in life processes was indicated in its derivative, vitamin B_2 (riboflavin, 6,7-dimethyl-9-(1'-D-ribityl isoalloxazine). Later, in the midsixties, it was identified in foods and its contributions to the unique flavor and aroma of raw and processed foods attracted the attention of flavor chemists Pyrazine derivatives contribute to the roasting, toasting, nutty, chocolaty, coffee, earthy, caramel, maple-like, bread-like, and bell pepper notes in foods. The reader is referred to the reviews on Krems and Spoerri ($\underline{88}$) on the chemistry of pyrazines, and the review of pyrazines in foods by Maga and Sizer ($\underline{89}$, $\underline{90}$). Table XVI summaries sensory description and threshold of selected pyrazines.

Oxygen Heterocyclics. During heat processing, sugars degrade to aldehydes and ketones which might react with amino compounds forming caramelized sugar flavors. Cyclic diketones, pyrones, and furan derivatives are examples of the products of this reaction. Table XVII gives the organoleptic description of selected compounds. 4-Hydroxy-2,5-dimethy1(2H)-furanone has intense fragrant caramel note described as burnt sweet taste (70), burnt pineapple $(\underline{101})$, beef broth $(\underline{100})$, strawberry preserve $(\underline{103})$, nutty sweet aroma of almonds (104), and major contributor to sponge cake flavor (105). Seven terms were used to describe the organoleptic note of one compound. The question arises, what was the concentration and media used for evaluation? Change in concentration alone can be quite sufficient to alter the character of the flavor or odor note. For example, trimethylamine-air mixtures only smell fishy over a narrow range of dilutions (1:1,500-1:8,000), with a maximum at about (1:6,000), also ammonia at a dilution of about (1:2,000) smells fishy ($\underline{106}$). Hodge and Moser ($\underline{107}$) reported that inspite of the fact that the aromas of pure maltol, ethyl maltol, isomaltol, and 4-hydroxy-2,5-dimethyl-3(2H)-furanone vary significantly from each other, panelists description was caramel or burnt. sugar. The fruity caramel aroma of isomaltol is weaker than that of 4-hydroxy-2,5-dimethyl-3(2H)-furanone (108). Our sensory vocabulary should be adequate to express the impact of flavor, odor, taste notes components. This could be achieved by the participation of food chemist, organic chemist, sensory analyst and flavorist. Soy sauce (Shoyu) contains tautomers 4-hydroxy-2-ethyl-5ethyl-3(2H)-furanone and 4-hydroxy-5-ethyl-2-methyl-3(2H)-furanone (about 3:2 ratio) which has sweet odor similar to that of short cake (109). The same tautomers were synthesized and described as possessing the flavor of cooked fruits (103). Maltol, isomaltol and 2-methyl-5-hydroxy-6-ethyl-α-pyrone are contributors to the characteristic aroma of molasses (110).

Table XVI

Pyrazines, Sensory Description and Odor Threshold

Pyrazine	Sensory Description	Water	Odor Threshold ppm Mineral Oil Vegetal	old ppm Vegetable Oil
2-Methy1-	deep bitter persistent cocoa	105 (91)	27 (91)	7000
2,3-Dimethyl-	notes (5)	(95)	\ \\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	
2,5-Dimethyl-	earthy raw potato (3)	2.5 (92) 3.5 (91)	7 (01)	
2,6-Dimethyl	sweet fried odor (2)	1.8 (92) 54 (91)	8 (01)	(76) 0.2
2,3,5-Tri-		1.5 (92)	(16) 0	
methyl		6 (61)	27 (91)	
2,3,5,6-Tetra-		(10) 01	000	
methyl o_r+x-		(TK) OT	36 (91)	
יי אוזאים ב		(26) 09	17 (91)	
2-Ethy1-3-		22 (91)		
methyl-		0.13 (92)		0.32 (92)
2-Ethyl-5- methyl		0.10 (92)		· ·
2-Ethy1-3,6-		(00)		
dimethyl-		(26) #000°0	1 1	0.024 (92)

Pable XVI

Pyrazines, Sensory Description and Odor Threshold (Cont.)

			Odor Threshold ppm	shold ppm
Pyrazine	Sensory Description	Water	Mineral Oil	Vegetable Oil
2 6-dimethown-3-	nuttv with oreen_like hell_			
isopropyl-5-methyl	pepper, woody notes (96)			
2,5-dimethoxy-3-	nutty notes, green (bell-			
isopropyl-6-methyl	pepper-like) (96)			
5-methyl 5H-6,7-	peanuts (53)	-		
dihydrocyclopenta				
5-methy1-2H-3,4,5,7-	roasted nuts, burnt (53)			
retrahydrocyclopenta				
2-acety1-	breadcrust, nutty (96)			
2-methoxy-3-acetyl-	weak breadcrust, nutty			
	notes (96)			
2-methylamino-3-	roasted peanuts, green cocoa			
methyl	notes (96)			
2-dimethylamino-3-	Peanut-like more green and			
methyl	less cocoa notes (96)			
2-dimethylamino-6-	peanut-like, no cocoa notes			
methyl	reminiscent of burnt coffee (96)	(9)		

Table XVI

Pyrasines, Sensory Description and Odor Threshold (Cont.)

Pyrazine	Sensory Description	Water	Mineral Oil	Mineral Oil Vegetable Oil
2,5-diethyl- 2,6-diethyl- 5-Ethyl-2,3- dimethyl	chocolate sweet (93)	0.02 (92)	-	0.27 (92)
2,5-Dimethyl-3- ethyl		43 (91)	24 (91)	
2,6-Dimethyl-3- ethyl		15 (91)	24 (91)	
Methyl isoamyl- Ethyl isoamyl- 2-Methylamine-3- methyl- 2-Dimethylamine-3- ethyl 2-Methyl-3-methoxy-	green cocoa note (95) green cocoa note (95) cocoa notes, roasted peanuts, green (96) less cocoa note (96)			
2-Methylthio-3-methyl-	roasted peanut (91) nutty earthy (97) nutty cracker (97)			

References (53, 91-97)

Table XVII
Oxygen Heterocyclic Compounds

Compound	Sensory Description
Furan	Spicy, smoky, slightly cinnamon- like odor (98).
	Sweet bread-like, caramel-like taste (98).
Furfuryl alcohol	Coconut (<u>99</u>).
5-methyl-2-furfural	Sharp grape (98) .
Maltol (3-hydroxy-2- methyl-pyrone)	Pleasing strong fruitsy fresh bread (99).
Isomaltol	Fruity caramel, fresh bread odor, overtone medicinal grassy (99) .
N-Furfuryl pyrrole	Green hay-like aroma (98) .
4-hydroxy-5-methyl- 3(2H)-Furanone	Beef broth $(\underline{100})$ burning sweet taste $(\underline{70})$, burnt pineapple $(\underline{101})$.
2,5-Dimethyl-4-hydroxy- 3(2H)-Furanone	Odor of roasted chicory roots (102) .

References (98 - 102).

Sulfur Heterocyclics. Sulfur containing compounds (thiols, thiophenes, thiazoles, ... etc.) play a major role in the flavor of raw and processed foods. These compounds have characteristic flavor notes and the flavor thresholds are mostly low. Several reviews (111, 112, 113) demonstrate the important role of sulfur compounds in food flavors. Organoleptic properties of these compounds may be pleasant, strong nut-like odor of 4-methyl-5-vinylthiazole which is present in cocoa (114); objectionable pyridine-like odor of thiazole (115); quinoline-like odor of benzothiazole (116); strong tomato leaf-like odor of isobutylthiazole (117); and bread crust flavor of acetyl-2-thiazoline (118). A mixture of oxazoles, thiazoles, thiazolines, imidazoles, trithiolanes and

dithianes which had a meaty flavor was obtained from a model system consisting of α -dicarbonyls, ethanal, hydrogen sulfide and ammonia (119). Unsaturated aldehydes react with hydrogen sulfide and thiols to give mainly addition products to the carbon-carbon double bond (120). The nomenclature of the resulting compounds and their organoleptic descriptions are given in Table XVIII. Thiols, thiophenes, thiazoles, sulfides and furans were identified in the volatiles of heating glucose, hydrogen sulfide, and ammonia at 100°C for two hours (121). These volatiles gave a roast beeflike aroma. Complex mixtures of mercapto-substituted furans and thiophene derivatives, which were reminiscent of roast meat were produced upon heating 4-hydroxy-5 methyl-3(2H)-furanone and its thio analog with hydrogen sulfide (122).

Lipid Browning.

Lipid browning reactions of the Maillard type between carbonyl groups (provided by sugars or sugar degradation products and those resulting from unsaturated fatty acids oxidation) and the free amino groups present in phospholipids have been recognized as potent causes of undesirable flavor, color and texture changes in dehydrated foods (123). Phosphatidyl ethanolamine, phoshatidyl serine, ethanolamine and serine plasmalogens contain free amino groups which can undergo lipid browning reactions. Phospolipids are usually rich in highly unsaturated fatty acids in comparison with neutral lipids, thus they are good sources of carbonyls. Also, the primary amine moities of polar lipids catalyze the aldol condensation of C_{14} - C_{18} aldehydes resulting from plasmalogen hydrolysis, thus forming α,β -unsaturated aldehydes ($\frac{124}{2}$). Phosphatidyl ethanolamine reacted with propanal and n-hexanal forming phosphatidyl 1-(2-hydroxyethyl)-2-ethyl-3,5-dimethyl pyridinium, and phosphatidyl-1-(2-hydroxyethyl)-2-hexyl-3,5-dipentyl pyridinium, respectively (125). The peridinium ring is formed by the reaction between one mole of amino-N of phosphatidyl ethanolamine and three moles of n-alkanals. The same reaction took place in the synthesis of substituted pyridines by condensation of carbonyl compounds with ammonia (126, 127).

Abstract

In processed foods, non-enzymic browning reaction is the major source of its desirable flavors. Flavors of the products of this reaction depend upon: the molecular structure of nitrogenous compounds (amines, amino acids, peptides, glycopeptides, proteins, ... etc.); aldoses, ketoses, non-reducing, deoxy sugars, sugar acids, ... etc.); heating temperature; duration of the reaction; initial hydrogen ion concentration, moisture content, and the media of the reaction (alcoholic, aqueous, or anhydrous). The ratio of the nitrogenous compound to sugar or carbonyl compound has great effect on the flavor notes. Comparison between browning

Table XVIII

Flavor of Products of the Addition Between Unsaturated Carbonyls and Methanethiol

Carbonyl Added	Addition Product	Flavor Impression	Water	Paraffin Oil	1
2-Butenal 2-Hexenal 2-Hebtenal 2-Nonenal 1-Octene-3-one	3-methyl thiobutanal 3-Methyl thiohexanal 3-Methyl thioheptanal 3-Methyl thiononal 1-Methyl thiooctan-3-one	Cheese-like (Brite) Cabbage (Rubbery) Unripe tomato Bast-like, slightly floral Radish-like	0, W W W	20 20 20 50 50	
Reference (120).					

reaction products and rate of degradation of reactants under anhydrous condition, aqueous and alcoholic media was discussed. Flavor notes of products of amino compounds-sugars reaction were reviewed with emphasis on amino-ribose system. Meat-like flavor was imparted by browning reaction products of carnosine-, citrulline-, histidine-, glutamic-, 2-pyrrolidone-5-carboxylic-, methionine-, cysteine-, cysteic-, and taurine-ribose. Recent advancements in nitrogen-, oxygen- and sulfur hetercyclics and lipid browning were presented.

Literature Cited.

- 1. Economics, Statistics, and Cooperative Service, USDA, National Review, 1978, April; p. 53.
- 2. Yamasaki, Y.; Kaekawa, K., Agr. Biol. Chem., 1978, 42, 1761.
- 3. Waley, S.G., Advan. Protein Chem., 1966, 21, 1.
- 4. Reineccius, G.A.; Andersen, D.A.; Kavanagh, T.E.; Keeney, P.G., J. Agr. Food Chem., 1972, 20, 1.
- 5. Patterson, R.L.S., J. Sc. Food Agr., 1968, 19, 31.
- 6. Wong, E.; Nixon, L.N.; Johnson, C.B., J. Agr. Food Chem., 1975, 23, 495.
- 7. Watanabe, K.; Sato, Y., Agr. Biol. Chem., 1970, 34, 88.
- 8. Watanabe, K.; Sato, Y., Agr. Biol. Chem., 1971, 35, 756.
- Yamato, T.; Tadao, K.; Kato, H.; Fujimaki, M., <u>Agr. Biol</u>. <u>Chem.</u>, 1970, <u>34</u>, 88.
- Hornstein, J., Chemistry and PhysTology of Flavors, Schultz, H.W.; Day, E.A.; Libbey, L.M., Eds., Avi Publishing Co., Westport, CT, 1967; pp. 228-250.
- 11. Watanabe, K.; Sato, Y., Agr. Biol. Chem., 1970, 34, 467.
- 12. Watanabe, K.; Sato, Y., Agr. Biol. Chem., 1971, 35, 278.
- Harkes, P.D.; Begemann, W.J., J. Am. Oil Chemists' Soc., 1974, 51, 356.
- 14. Hornstein, I.; Crowe, P.F.; Heimberg, M.J., <u>J. Food Sci.</u>, 1961, <u>26</u>, 581.
- 15. Pippen, E.L.; Nonaka, M.; Jones, F.T.; Stitt, F., Food Res., 1958, 23, 103.
- 16. Pippen, E.L.; Nonaka, M., Food Res., 1960, 25, 764.
- 17. Heyns, K.; Stute, R.; Paulsen, H., <u>Carbohydrate Res.</u>, 1966, 2, 132.
- 18. Heyns, K.; Klier, M., Carbohydrate Res., 1968, 6, 436.
- 19. Walter, R.; Fagerson, I.S., <u>J. Food Sci.</u>, 1968, <u>33</u>, 294.
- 20. Kort, M.J., Advan. Carbohydrate Chem. Biochem., 1970, 25, 311.
- 21. Chuyen, N.V.; Kurata, T.; Fujimaki, M., Agr. Biol. Chem., 1973, 37, 327.
- 22. Kato, S.; Kurata, T.; Fujimaki, M., Agr. Biol. Chem., 1973, 37, 1759.
- 23. Fujimaki, M.; Kato, S.; Kurata, T., <u>Agr. Biol. Chem.</u>, 1969, 33, 1144.
- 24. Kato, S.; Kurata, T.; Fujimaki, M., Agr. Biol. Chem., 1973, 37, 539.

- Schonberg, A.; Moubacher, R., Chem. Rev., 1952, 50, 261.
- Hodge, J.E., J. Agr. Food Chem., 1953, 1, 928.
- 27. Hodge, J.E., Advan. Carbohydrate Chem., 1955, 10, 169.
- Ellis, G.P., Advan. Carbohydrate Chem., 1959, 14, 163.
- Reynolds, T.M., Advan. Food Res., 1963, 12, 1.
- Reynolds, T.M., Advan. Food Res., 1965, 14, 167.
- Lightbody, H.D.; Feuold, H.R., Advan. Food Res., 1948, 1, 149.
- 32. Ross, A.F., Advan. Food Res., 1948, 1, 325.
- Stadman, E.R., Advan. Food Res., 1948, 1, pp. 325-372. 33.
- 34. Wodemeyer, G.A.; Dollar, A.M., J. Food Sci., 1963, 28, 537.
- Wiseblatt, L.; Zoumut, H.F., Cereal Chem., 1963, 40, 162. 35.
- 36. El'Ode, K.E.; Dornseifer, T.P.; Keith, E.S.; Powers, J.J., J. Food Sci., 1966, 31, 351.
- 37. Herz, W.J.; Shallenberger, R.S., Food Res., 1960, 25, 491.
- Barnes, H.M.; Kaufman, C.W., Ind. Eng. Chem., 1947, 39, 1167. 38.
- Kiley, P.J.; Nowlin, A.C.; Mariarty, J.H., Cereal Sci. Today, 1960, 5, 273.
- Buckdeschel, W.Z., Ges Brau, 1914, 430, thru Chem. Abs., 40. 1915, <u>9</u>, 118.
- Morimoto, T.; Johnson, J.A., Cereal Chem., 1966, 43, 627.
- Keeney, M.; Day, E.A., <u>J. Dairy Sci.</u>, 1957, <u>40</u>, 847.
- Actander, S., Perfume and Flavor Chemicals (Aroma Chemicals); Published by the Author, Montclair, NJ, 1969; two volumes.
- 44. Persson, T.; von Sydow, E., <u>J. Food Sci.</u>, 1973, <u>38</u>, 377.
- Rooney, L.W.; Salem, A.; Johnson, J.A., Cereal Chem., 1967. <u>44</u>, 539.
- 46. Self, R., Chemistry and Physiology of Flavors, Schultz, H.W.; Day, E.A.; Libbey, L.M., Eds., Avi Publishing Co., Westport, CT, 1967; pp. 362-389.
- 47. Mabrouk, A.F.; Holms, L.G., Abstracts of Papers, 159, Division of Agricultural and Food Chemistry, 172nd ACS National Meeting, San Francisco, CA, September 1976.
- Rohan, T.A.; Stewart, T., J. Food Sci., 1966, 31, 202.
- Rohan, T.A.; Stewart, T., <u>J. Food Sci.</u>, 1966, <u>31</u>, 206.
- Rohan, T.A.; Stewart, T., <u>J. Food Sci.</u>, 1967, <u>32</u>, 395. Rohan, T.A.; Stewart, T., <u>J. Food Sci.</u>, 1967, <u>32</u>, 399.
- Rohan, T.A.; Stewart, T., J. Food Sci., 1967, 32, 625. 52.
- van Praag, M.; Stein, S.H.; Tibbets, M.S., J. Agr. Food Chem., 53. 1968, 16, 1005.
- Pickenhagen, W.; Dietrich, P.; Keil, B.; Polansky, J.; Nouaille, F.; Lederer, E., Helv. Chem. Acta., 1975, 58, 1078.
- Kato, S.; Kurata, T.; Ishitsuka, R.; Fujimaki, M., Agr. Biol. 55. Chem., 1970, 34, 1826.
- Shiba, T.; Nunami, K.I., Tetrahedron Letters, 1974, 6, 509.
- Takanishi, K.; Tadenuma, M.; Kitamoto, K.; Sato, S., Agr. Biol. Chem., 1974, <u>38</u>, 927.
- Sakamura, S.; Furukawa, K.; Kasai, T., Agri. Biol. Chem., 1978, 42, 607.
- Boyd, E.N.; Keeney, P.G.; Patton, S., J. Food Sci., 1965, 30, 59+

- 60. van Straten, S.; de Vrijer, F., <u>List of Volatile Compounds in Food</u>, Central Institute for Nutrition and Food Research, Zeist, Holland, 1973.
- 61. Eckey, E.W., <u>Vegetable Fats and Oils</u>, Reinhold Publishing Corp., NY, 1954; p. 676, 760.
- Feldman, J.R.; Ryder, W.S.; Kung, J.T., <u>J. Agr. Food Chem.</u>, 1969, <u>17</u>, 733.
- 63. Underwood, G.E.; Deatherage, F.E., Food Res., 1952, 17, 425.
- 64. Khan, N.A.; Brown, J.B., <u>J. Am. Oil Chemists' Soc.</u>, 1953, 30 606.
- 65. Shigematsu, H.; Kurata, T.; Kato, H.; Fujimaki, M., Agr. Biol. Chem., 1972, 36, 1631.
- 66. Kobayasi, N.; Fujimaki, M., Agr. Biol. Chem., 1965, 29, 1059.
- 67. Yoshikawa, K.; Libbey, L.M.; Cobb, W.Y.; Day, E.A., <u>J. Food</u> Sci., 1965, <u>30</u>, 991.
- Gianturco, M.A.; Giammarino, A.S.; Friedel, P.; Flanagan, V., <u>Tetrahedron</u>, 1964, 20, 2951.
- Gianturco, M.A.; Giammarino, A.S.; Friedel, P., <u>Nature</u>, 1966, <u>210</u>, 1358.
- 70. Tatum, J.H.; Shaw, P.E.; Berry, R.E., <u>J. Agr. Food Chem.</u>, 1967, <u>15</u>, 773.
- 71. Kato, H., Agr. Biol. Chem., 1966, 30, 822.
- 72. Kato, H., Agr. Biol. Chem., 1967, 31, 1086.
- 73. Kato, H.; Fujimaki, M., J. Food Sci., 1968, 33, 445.
- 74. Kato, H.; Fujimaki, M., Agr. Biol. Chem., 1970, 34, 1071.
- 75. Shigematsu, H.; Kurata, T.; Kato, H.; Fujimaki, M., Agr. Biol. Chem., 1971, 35, 2097.
- Kato, H.; Sonobe, H.; Fujimaki, M., <u>Agr. Biol. Chem.</u>, 1977, 41, 711.
- 77. Sonobe, H.; Kato, H.; Fujimaki, M., Agr. Biol. Chem., 1977, 41, 609.
- 78. Rizzi, G.P., J. Agr. Food Chem., 1974, 22, 279.
- 79. Shigematsu, H.; Kurata, T.; Kato, H.; Fujimaki, M., Agr. Biol. Chem., 1972, 36, 1631.
- 80. Walradt, J.P.; Pittet, A.O.; Kinlin, T.E.; Muralidhara, R.; Sanderson, A., J. Agr. Food Chem., 1971, 19, 972.
- 81. Kinlin, T.E.; Muralidhara, R.; Pittet, A.O.; Sanderson, A.; Walradt, T.P., J. Agr. Food Chem., 1972, 20, 1021.
- 82. Hunter, I.R.; Walden, M.K.; Scherer, J.R.; Lundin, R.E., Cereal Chem., 1969, 46, 189.
- 83. Watanabe, K.; Sato, Y., <u>J. Agr. Food Chem.</u>, 1971, <u>19</u>, 1017.
- 84. Buttery, R.G.; Ling, L.C.; Teranishi, R.; Mont, T.R., <u>J. Agr. Food Chem.</u>, 1977, <u>25</u>, 1277.
- 85. MacLeod, G.; Coppock, B.M., J. Agr. Food Chem., 1977, 25, 113.
- 86. Webster's Third New International Dictionary, G&C Merriam Co., Publishers, Springfield, MA, 1967.
- Goldman, I.M.; Seibl, J.; Flament, I.; Gautschi, F.; Winter,
 M.; Willham, B.; Stoll, M., Helv. Chem. Acta., 1967, 50, 694.
- 88. Krems, I.J.; Spoerri, P.E., Chem. Rev., 1947, 40, 279.

- Maga, J.A.; Sizer, C.E., Crit. Rev. Food Technol., 1973, 4, 89.
- Maga, J.A.; Sizer. C.E., J. Agr. Food Chem., 1973, 21, 22.
- Seifert, R.M.; Buttery, R.G.; Guadagni, D.G.; Black, D.R.; Harris, J.G., <u>J. Agr. Food Chem.</u>, 1970, <u>18</u>, 246.
- Seifert, R.M.; Buttery, R.G.; Guadagni, D.G.; Black, D.R.; Harris, J.G., <u>J. Agr. Food Chem.</u>, 1972, <u>20</u>, 135.
- Deck, R.E.; Chang, S.S., Chem. Ind., 1965, 1343.
- Buttery, R.G.; Seifert, R.M.; Guadagni, D.G.; Ling, L.C., J. Agr. Food Chem., 1969, 17, 1322.
- Polak's Fruital Works, Improvements relating to flavorings, Br. Patent 1,248,380, September 29, 1971.
- Takken, H.J.; van der Linde, L.M.; Boelens, M.; van Dort, J. 96. M., J. Agr. Food Chem., 1975, 23, 638.
- Parliment, T.H.; Epstein, M., J. Agr. Food Chem., 1973, 21, 714.
- Guadagni, D.G.; Buttery, R.G., J. Sci. Food Agr., 1972, 23, 1435.
- Shaw, P.E.; Tatum, J.H.; Berry, R.E., J. Agr. Food Chem., 1969, <u>17</u>, 907.
- 100. Tonsbeek, C.H.T.; Planchen, A.J.; van de Weerdhof, T., J. Agr. Food Chem., 1968, 16, 1016.
- 101. Rodin, J.O.; Himel, C.M.; Silverstein, R.M.; Leeper, R.W.; Gortner, W.A., <u>J. Food Sci.</u>, 1965, <u>30</u>, 280.
- 102. Tonsbeek, C.H.T.; Koenders, E.B.; van der Zijden, A.S.M.; Losekoot, J.A., <u>J. Agr. Food Chem</u>., 1969, <u>17</u>, 397.
- 103. Re, V.L.; Maurer, B.; Ohloff, G., Helv. Chem. Acta., 1973, <u>56</u>, 1882.
- 104. Takei, Y.; Yamanishi, T., Agr. Biol. Chem., 1974, 38, 2329.
- 105. Takei, Y., Agr. Biol. Chem., 1977, 41, 2361.
- 106. Stansby, M.E., Food Technol., 1962, 16, 28.
- 107. Hodge, J.E.; Moser, H.A., Cereal Chem., 1961, 38, 221.
- 108. Hodge, J.E.; Mills, F.D.; Fisher, B.E., Cereal Sci. Today, 1972, <u>17</u>, 34.
- 109. Numomura, N.; Saaki, M.; Asao, Y.; Yokotsuka, T., Agr. Biol. Chem., 1976, 40, 491.
- 110. Ito, H., Agr. Biol. Chem., 1976, 40, 827.
- 111. Schutte, L., Crit. Rev. Food Technol., 1974, 4, 457.
- 112. Maga, J.A., Crit. Rev. Food Sci. Nutr., 1975, 6, 153.
- 113. Maga, J.A., Crit. Rev. Food Sci. Nutr., 1975, 6, 241.
- 114. Maga, J.A., Crit. Rev. Food Sci. Nutr., 1976, 7, 147.
- 115. Stoll, M.; Dietrich, P.; Sunte, E.; Winter, M., Helv. Chem. Acta., 1967, 50, 2065.
- 116. Merck Index 9th ed., 1976; p. 1313.
- 117. Viani, R.; Bricout, J.; Marion, J.P.; Muggler-Chavan, F.; Reymond, D.; Egli, R.H., Helv. Chem. Acta., 1969, 52, 887.
- 118. Tonsbeek, C.H.T.; Copier, H.; Plancken, A.J., J. Agr. Food Chem., 1971, 19, 1014.

- Boelens, M.; van der Linde, L.M.; de Valois, P.J.; van Dort, H.M.; Takken, H.J., Proc. Int. Symp. Aroma Research, Zeist, Netherland, 1975; pp. 95-100.
- 120. Badings, H.T.; Maarse, H.; Kleipool, R.J.C.; Tas, A.C.; Neeter, R.; ten Noever de Brauw, M.C., Proc. Int. Symp. Aroma Research, Zeist, Netherland, 1975; pp. 63-73.
- 121. Shibamoto, T.; Russell, G.F., J. Agr. Food Chem., 1976, 24,
- 122. van den Ouweland, G.A.M.; Peer, H.G., J. Agr. Food Chem., 1975, <u>23</u>, 501.
- 123.
- Lea, C.H., J. Sci. Food Agr, 1957, 8, 1.
 Nakanishi, T.; Suyama, K., Nippon Chikusan Gakkai-Ho, 1970, 124.
- 125. Nakanishi, T.; Suyama, K., <u>Agr. Biol. Chem</u>., 1974, <u>38</u>, 1141.
- Frank, R.L.; Seven, R.P., J. Amer. Chem. Soc., 1949, 71,
- Farley, C.P.; Eliel, E.L., J. Amer. Chem. Soc., 1956, 78, 127. 3477.

RECEIVED August 2, 1979.